ORIGINAL ARTICLE

Analgesic and anti-inflammatory studies of cyclopeptide alkaloid fraction of leaves of *Ziziyphus nummularia*

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KEYWORDS

*Ziziyphus nummularia*; Analgesic; Anti-inflammatory; Cyclopeptide alkaloids; Cotton pellet granuloma; Acetic acid induced writhing; Tail flick; Hot plate

Abstract

*Ziziyphus nummularia* (family: Rhamnaceae) is a thorny small bush, grows in abundance in the grazing lands of the arid areas of Rajasthan, India. It is an important ethnomedicinal plant of the Thar Desert; local inhabitants use every part of the plant as medicine. Kernels are prescribed in pregnancy as soporific, antiemetic and for relieving abdominal pain. The insect gall is powdered and given orally with water to cure bone fracture. Crushed root is applied on the paining shoulder of the bullock. The decoction of leaves is used for the treatment of cough and cold; leaves are also regarded as diaphoretic and prescribed in typhoid. Paste of leaves is used for healing of cuts, boils and cutaneous disease. It is widely used in pain and inflammatory conditions.

*Z. nummularia* contains a unique group of alkaloids known as cyclopeptide alkaloids, in continuation of our work carried out on the leaves of *Z. nummularia*, present study was initiated to explore anti-inflammatory and analgesic potential of cyclopeptide alkaloids isolated from the leaves of *Z. nummularia* (IFZN). Anti-inflammatory activity was tested against rat paw oedema, mouse peritonitis and cotton pellet granuloma. For screening of analgesic activity, acetic acid induced writhing, tail flick and hot plate test were performed.

IFZN 30 mg/kg shows the anti-oedematogenic effect against paw oedema induced by carrageenan, dextran, serotonin and histamine; IFZN 20 and 30 mg/kg were found to have highly significant anti-nociceptive effects.

Result of pharmacological studies indicated that IFZN is a potent and efficacious analgesic agent. The analgesic activity of IFZN is mediated by the peripheral as well as central pathways.

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1. Introduction

*Ziziyphus nummularia* (family: Rhamnaceae) is a thorny small bush, grows in abundance in the grazing lands of the arid areas of Rajasthan, India (Anonymous, 1989). It is an important
ethnomedicinal plant of the Thar Desert; local inhabitants use every part of the plant as medicine. Kernels are prescribed in pregnancy as a soporific, antieptic and for relieving abdominal pain (Singh and Pandey, 1998). Filtrate of fruit decoction is used for bathing to cure fever caused due to heat stroke (Kumar et al., 2008; Meena and Yadav, 2010). Extract of fruit is also used for the treatment of infertility (Muhammad and Khan, 2008).

Root is useful in the treatment of diarrhoea and dysentery (Jadhav, 2006). The young branches and roots are used as toothbrushes for preventing decay, pyorrhea and other diseases of the teeth (Jain et al., 2005; Upadhya et al., 2011). The insect gall is powdered and given orally with water to cure bone fracture. Crushed root is applied on the paining shoulder of the bullock, used in ploughing (Bhattacharjee, 2004; Jadeja et al., 2006).

The decoction of leaves of *Z. nummularia* is used for the treatment of cough and cold, leaves are regarded as diaphoretic and prescribed in typhoid. Paste of leaves is used for healing of cuts, boils and cutaneous disease (Meena and Yadav, 2010). It is also used in pain and inflammatory conditions (Kumar et al., 2008; Goyal et al., 2011).

We have reported anti-inflammatory and analgesic effects of an ethanolic extract of leaves of *Z. nummularia* (EZN) against rat paw oedema, mouse peritonitis, acetic acid induced writhing and tail flick test, respectively (Goyal et al., 2012). Similarly, alcoholic extract of leaves has also been reported for topical anti-inflammatory and wound healing effects (Hasan Soliman Yusufoglu, 2011).

Phytochemical analysis of *Z. nummularia* revealed the presence of number of phytoconstituents such as flavonoids, tannins, sterols, saponins, pectin, glycosides, alkaloids and triterpenoic acids. However, cyclopeptide alkaloids are regarded as active constituents responsible for pharmacological actions of *Z. nummularia* (Morel Ademir et al., 2009; Hasan Soliman Yusufoglu, 2011; Goyal et al., 2012). Therefore, in continuation of our work carried out on leaves of *Z. nummularia*, present study was initiated to explore the anti-inflammatory and analgesic potential of cyclopeptide alkaloids isolated from the leaves of *Z. nummularia* (IFZN). Receptor docking studies were also conducted in addition to *in vivo* pharmacological studies to understand the possible mechanism of action of IFZN.

2. Materials and methods

2.1. Plant material

The leaves of *Z. nummularia* were collected from Jodhpur, Rajasthan. Plant was identified by Taxonomists of the Botanical Survey of India (BSI), Jodhpur, and a voucher specimen BSI/AZC/MG-1 was deposited for future reference.

2.2. Extraction of IFZN

IFZN was isolated as per the method described by Ma et al. (2008). One kilogram of dried and powdered leaves of *Z. nummularia* was extracted in a soxhlet extractor for 24 h with hexane. The residue was extracted with ethanol for 24 h and evaporated to dryness by rotary evaporation. The obtained ethanolic extract was partitioned between 5% hydrochloric acid (60 ml) and ether (50 ml). The aqueous acid solution was extracted further three times with chloroform (60 ml · 3) in the presence of ammonia hydrate (quantity sufficient to produce pH 9.0). The cyclopeptide alkaloid fractions in the chloroform solution were combined, filtered through Whatman No. 1 filter paper, and concentrated using a rotary vacuum evaporator (Ma et al., 2008). The yield of IFZN was found to be 0.02%. The presence of a cyclopeptide alkaloid was confirmed by identification tests, TLC and GCMS analysis.

2.3. Animals

Healthy Wistar rats of both sexes weighing 180–200 g and Swiss mice weighing 20–24 g were used. The animals had free access to a standard commercial diet and water ad libitum and were kept in rooms maintained at 25 °C in a 12-h light/
dark cycle. The animals were used after an acclimatization period of 7 days to the laboratory environment. The experiments were performed during the light portion (0800–1600 h). The experimental protocol and procedures used in this study were approved by IAEC of the Lachoo Memorial College of Science and Technology, Pharmacy Wing, Jodhpur vide the approval number LMC/OFFICE/2008/IAEC/17/5/2008/PHD/2.

2.4. Chemicals and drugs

Carrageenan (S.D. Fine Chemicals Limited, Bombay), histamine, serotonin, dextran (Sigma, USA), indomethacin (Recon, Bangalore), aspirin (USV Bombay) and morphine (Bio E) were used in the study.

2.5. Acute toxicity study

Mice were kept on overnight fasting and water was withheld for 3–4 h before the administration of the test compound. Following the period of fasting, the animals were weighed and divided into five groups, three mice in each group. The control group received normal saline 1 ml/kg BW by gavage while the treatment groups received IFZN in a dose of 5 mg/kg BW. Immediately after dosing, the mice were observed continuously for 4 h for symptoms of toxicity like motor activity, tremors, convulsions, tonic extension, muscle spasm, loss of righting reflex, ataxia, sedation, hypnosis, lacrimation, diarrhea, salivation and writhing. Mice were then kept under observation up to 48 h for any mortality, next higher dose 50 mg/kg and 100 mg/kg and IFZN at doses of 10, 20, 30 mg/kg BW per oral for seven consecutive days from the day of cotton pellet implantation. The mice were anaesthetized on the eighth day and cotton pellets were removed surgically (Penn and Ashford, 1963). Pellets were made free from extraneous tissues, moist pellets were weighed and then dried at 60 °C for 24 h, after that dried pellets were weighed again. Increment in the weight of the pellets was taken as a measure of granuloma formation.

2.6. Anti-inflammatory activity

2.6.1. Carrageenan-induced rat paw oedema test

Carrageenan-induced rat paw oedema is the most commonly used technique for the screening of anti-inflammatory drugs. The test is based upon the ability of the drug to inhibit the acute edema produced in the hind paw of the rat after injection of a phlogistic agent such as carrageenan, dextran, histamine or serotonin.

Five groups of rats, six in each group were selected for the study. Different groups were treated with saline; indomethacin 10 mg/kg BW and IFZN at doses of 10, 20, 30 mg/kg BW per oral. The animals were treated with the extract 1.0 h before the administration of carrageenan. Acute inflammation was produced by the subplantar administration of 0.1 ml of 1% carrageenan in normal saline in the right hind paw of the rats. The paw volume was measured at 0, 1.0 and 3.0 h after carrageenan injection using plethysmometer (Winter and Porter, 1957). The anti-inflammatory effect was calculated by the following equation:

\[
\text{Anti-inflammatory activity} \% = \left(1 - \frac{D}{C}\right) \times 100.
\]

Where D represents the percentage difference of the paw volume of treated groups and C represents the percentage difference of volume in the control group (Suleyman et al., 1991).

2.6.2. Dextran, histamine and serotonin-induced paw oedema

The rats were treated in a manner similar to that of carrageenan-induced paw oedema models; dextran (0.1 ml, 1% w/v in normal saline), 0.1 ml of freshly prepared histamine (1 mg/kg BW) and serotonin (1 mg/kg BW) were used in the place of carrageenan (Winter and Porter, 1957; Winter et al., 1962).

2.6.3. Cotton pellets-induced granuloma

Cotton pellets induced granuloma are used for the evaluation of steroidal and nonsteroidal anti-inflammatory drugs. The amount of newly formed connective tissue (granuloma) can be measured by this method.

Five groups of mice, six in each group were selected for the study. Mice were anesthetized and fur of their back was removed with an electric razor. Subsequently, an incision was made in the lumbar region and with the help of a blunted forceps; subcutaneous tunnel was made on right side in the scapular region. Sterilized cotton pellet weighing 15 mg was placed in the pouch formed in the scapular region. The different groups of mice were treated with saline; indomethacin 10 mg/kg and IFZN at doses of 10, 20, 30 mg/kg BW per oral for seven consecutive days from the day of cotton pellet implantation. The mice were anaesthetized on the eighth day and cotton pellets were removed surgically (Penn and Ashford, 1963). Pellets were made free from extraneous tissues, moist pellets were weighed and then dried at 60 °C for 24 h, after that dried pellets were weighed again. Increment in the weight of the pellets was taken as a measure of granuloma formation.

2.6.4. Mouse carrageenan peritonitis

Carrageenan induced peritonitis in mice is used for accessing the effect of anti-inflammatory drugs on fluid extravasation and leukocyte migration induced due to inflammation (Grishwold et al., 1987).

Five groups of mice, six in each group were selected for the study. Different groups of mice were treated with saline; indomethacin 10 mg/kg BW and IFZN at doses of 10, 20, 30 mg/kg BW per oral. One hour later peritonitis was induced by intraperitoneal injection of carrageenan (0.25 ml, 0.75% w/v in saline). After 4 h, the animals were sacrificed and peritoneal fluid collected for leukocyte count (Griswold et al., 1987).

2.7. Analgesic activity

Pain is frequently associated with inflammation; it is the final product of a complex information processing network involving the central and peripheral pathways. Drugs with anti-inflammatory effect very often possess analgesic properties as well. Analgesic activity of IFZN was evaluated against acetic acid-induced writhing response, which is a method for measuring peripheral analgesic activity. Central analgesic effect was measured by the tail flick test and hot plate.

2.7.1. Acetic acid-induced writhing response in mice

Five groups of mice, six in each group were selected for the study. The different groups were treated with saline; aspirin 100 mg/kg and IFZN at doses of 10, 20, 30 mg/kg BW per oral. Thirty minutes after the drug treatment, 0.1 ml of a 0.6% solution of acetic acid was injected intraperitoneally
and the number of writhes during the following 30 min were counted (Koster et al., 1959). Percentage inhibition was calculated by the following equation: Percentage inhibition = \((1 - D/C) \times 100\)

Where \(D\) represents the number of writhes of treated groups and \(C\) represents the number of writhes of the control group.

2.7.2. Tail flick test

The tail flick test in mice is used for screening of centrally acting analgesic drugs. Mice are placed into cages leaving the proximal third of the tail exposed to the radiant heat source. Within a few seconds the animal flicks the tail aside or tries to escape. The time until this reaction occurs is measured.

Six groups of mice, six in each group were selected for the study. Mice which had 3.5–4.5 s baseline latency of tail flick were included in study. First four groups were treated with saline and IFZN at doses of 10, 20, 30 mg/kg BW per oral, fifth group was treated with a subcutaneous injection of morphine 5 mg/kg BW and the sixth group was treated with naloxone (1 mg/kg BW i.p.) followed by IFZN 30 mg/kg BW per oral. IFZN, morphine and naloxone were given 60, 30 and 15 min before the test, respectively. Mice were screened by placing their proximal third of the tail to the radiant heat source maintained at 50 ± 2°C, latency for tail flick was recorded. A cutoff time of 15 s was used to avoid damage to the tail (D’Armour and Smith, 1941).

2.7.3. Hot plate test

Hot plate test is specifically used for the screening of centrally acting analgesics. Administration of centrally acting analgesics causes prolongation of the latency times of response i.e. jumping, withdrawal of the paws and licking of the paws.

Five groups of mice, six in each group were selected for the study. The mice which reacted within 15 s and which did not show a large variation when tested on two separate occasions were included in study. First four groups were treated with saline and IFZN at doses of 10, 20, 30 mg/kg BW per oral, fifth group treated with subcutaneous injection of morphine 5 mg/kg BW. IFZN and morphine were given 60 and 30 min before the test, respectively. Mice were screened by placing them on a hot plate maintained at 55 ± 2°C and recorded the reaction time in seconds for the licking of paw or jumping (Turner, 1965). A cutoff time of 15 s was used to avoid damage to the paw.

2.8. Docking study

Docking studies were performed against X-ray crystal structure of the \(\mu\)-opioid receptor (2IQO) using LeadIT FlexX Version: 1.0.1 (BiosolveIT GmbH, Sankt Augustin, Germany).

Structures of ligands (Morphine, Nummularine and Ziziphus nummularia) were drawn using Marvin Sketch-5.0.6.1 (ChemAxon Ltd.). The 3D-geometry optimized ligands were saved in .sdf format. Crystal structure of the \(\mu\)-opioid receptor (2IQO) was downloaded from the Protein data bank (Rscb, 2012).

Docking protocol: Triangle matching base fragment placement and all the poses were used for docking. 200 solutions per iteration and 200 solutions per fragmentation were used.

2.9. Statistical analysis

Results were statistically analysed by ANOVA, followed by the Student’s \(t\)-test. The results were expressed as mean ± standard error mean (SEM). \(P < 0.05\) was considered significant.

3. Results

3.1. Acute toxicity study

According to OECD guidelines, IFZN falls in category-3 having LD\(_{50}\) 200 mg/kg.

Body weight, RBC and WBC count, and weight of heart and liver of treated mice were found to be well within the limit. However, CNS depression and reduction in locomotor activity were observed in mice treated with IFZN 200 and 300 mg/kg.

Therapeutic range was considered between 1/20 and 1/5 times of LD\(_{50}\). Accordingly, 10, 20 and 30 mg/kg BW of IFZN were selected as doses for pharmacological studies.

3.2. Anti-inflammatory activity

IFZN shows the dose-dependent anti-oedematogenic effects against paw oedema induced by carrageenan, dextran, serotonin and histamine Fig. 1. Results of the paw oedema test are summarized in Table 1.

Carrageenan induced peritonitis in mice was used to access fluid extravasation and leukocyte migration induced due to inflammation. IFZN causes significant dose dependent

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Dose mg/kg BW (p.o.)</th>
<th>Increase in paw oedema in milliliters (mean ± SEM)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Carrageenan 1h 3h</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Dextran 1h 3h</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Histamine 1h 3h</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Serotonin 1h 3h</td>
</tr>
<tr>
<td>Control</td>
<td>5 (ml/kg)</td>
<td>0.90 ± 0.02 0.94 ± 0.03</td>
</tr>
<tr>
<td></td>
<td></td>
<td>0.92 ± 0.02 0.98 ± 0.05</td>
</tr>
<tr>
<td></td>
<td></td>
<td>0.94 ± 0.01 1.02 ± 0.05</td>
</tr>
<tr>
<td></td>
<td></td>
<td>0.95 ± 0.01 0.99 ± 0.03</td>
</tr>
<tr>
<td>Indomethacin</td>
<td>10</td>
<td>0.36 ± 0.03 0.25 ± 0.03(^e)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>0.41 ± 0.02 0.25 ± 0.04(^e)</td>
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<tr>
<td></td>
<td></td>
<td>0.36 ± 0.01 0.25 ± 0.01(^e)</td>
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<tr>
<td></td>
<td></td>
<td>0.42 ± 0.01 0.25 ± 0.01(^e)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>0.25 ± 0.01(^e)</td>
</tr>
<tr>
<td>IFZN I</td>
<td>10</td>
<td>0.57 ± 0.02 0.43 ± 0.05(^e)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>0.62 ± 0.01 0.46 ± 0.02(^e)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>0.53 ± 0.01 0.48 ± 0.02(^e)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>0.63 ± 0.01 0.48 ± 0.02(^e)</td>
</tr>
<tr>
<td>IFZN II</td>
<td>20</td>
<td>0.50 ± 0.02 0.40 ± 0.04(^e)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>0.51 ± 0.01 0.43 ± 0.03(^e)</td>
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<tr>
<td></td>
<td></td>
<td>0.50 ± 0.01 0.48 ± 0.03(^e)</td>
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<tr>
<td></td>
<td></td>
<td>0.56 ± 0.02 0.46 ± 0.03(^e)</td>
</tr>
<tr>
<td>IFZN III</td>
<td>30</td>
<td>0.47 ± 0.01(^e) 0.38 ± 0.04(^e)</td>
</tr>
<tr>
<td></td>
<td></td>
<td>0.46 ± 0.01(^e) 0.41 ± 0.02(^e)</td>
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<td>0.44 ± 0.01(^e) 0.42 ± 0.02(^e)</td>
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<td></td>
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<td>0.52 ± 0.03(^e) 0.44 ± 0.04(^e)</td>
</tr>
</tbody>
</table>

Values reported as mean ± SEM (n = 6). The data were analyzed by one way ANOVA followed by Dunnett’s test.

* \(P < 0.05\) as compared with the control group. IFZN (cyclopeptide alkaloid fraction of leaves of Ziziphus nummularia).
decreases in the number of leukocytes migrating to the peritoneal cavity (Table 2).

Effects of IFZN treatment in animals implanted with cotton pellet-induced granuloma are depicted in Table 2. IFZN treatment causes significant \((P < 0.05)\) dose dependent decreases in the weight of moist and dry cotton pellets in comparison to the control group.

### 3.3. Analgesic activity

The analgesic effects induced by different doses of IFZN and aspirin 100 mg/kg BW on the acetic acid induced writhing in mice are portrayed in Fig. 2. Significant \((P < 0.05)\) dose dependent decrease in the number of writhes was observed in IFZN treated animals. The results obtained for the writhing test are similar to those obtained for the oedematogenic test using carrageenan.

Results of tail flick and hot plate tests are summarized in Table 3. IFZN and morphine 5 mg/kg causes significant \((P < 0.05)\) dose dependent increases in tail flick latency and hot plate reaction time when compared with the control. Co-administration of naloxone causes moderate decrease in the tail flick latency of mice when compared with IFZN 30 mg/kg.

### 3.4. Docking studies

The docking scores nummularine on \(\mu\)-opioid were found to be \(-14.00\). Docking analysis of docked conformers on \(\mu\)-opioid receptor revealed that nummularine interacts with PHE 237 and TRP 303. Nummularine has also displayed H-bonding interaction with THR 307 (2.02 \(\AA\)) and LYS 303 (1.87 \(\AA\)).

### 4. Discussion

In comparison to our earlier study carried out with alcoholic extracts of leaves of ZN, IFZN was found to be an efficacious and potent analgesic agent.

![Figure 2](image.png)  
**Figure 2** Effect of IFZN on acetic acid induced writhing in mice. Values reported as mean ± SEM \((n = 6)\). The data were analyzed by one way ANOVA followed by the Dunnett’s test. *\(P < 0.05\); **\(P < 0.01\) as compared with the control group. IFZN (cyclopeptide alkaloid fraction of leaves of *Ziziphus nummularia*).
Anti-inflammatory activity of IFZN is purposed to be mediated by the inhibition of prostaglandin synthesis. Reduction in paw oedema induced by carrageenan and histamine, decreased weight of moist cotton pellet and decreased leukocyte count in carrageenan induced peritonitis are the indicators of decreased vascular permeability and cellular migration to the injured site (Cuman et al., 2001; Linardi et al., 2002). These effects are known to be mediated by the release of histamine, serotonin, cytokines, leukotrienes and prostaglandins (Hwang et al., 1996).

Further, inhibition of the carrageenan induced neutrophil migration to the peritoneal cavity and decreases in acetic acid-induced writhing are indicators of inhibitory action on leukotriene and cytokine synthesis (Canetti et al., 2001; Limet and Lecomte, 1968). These effects are known to be mediated by the release of histamine, serotonin, cytokines, leukotrienes and prostaglandins (Hwang et al., 1996).

Further, inhibition of the carrageenan induced neutrophil migration to the peritoneal cavity and decreases in acetic acid-induced writhing are indicators of inhibitory action on leukotriene and cytokine synthesis (Canetti et al., 2001; Limet and Lecomte, 1968).

IFZN 30 mg/kg has shown highly significant anti-nociceptive effects against acetic acid-induced writhing, tail flick and hot plate test (Fig. 3).

Administration of acetic acid causes increased membrane phospholipase activity and synthesis of pain and inflammation mediators such as cytokines, eicosanoids, leukotrienes and prostaglandins; therefore protection against acetic acid induced writhing is indicative of the inhibition of prostaglandin like mediators (Eun-Mi Choi and Hwang, 2004).

The hot-plate paw-licking as well as the tail-flick responses are used for studying the neuronal mechanisms of opioid and stimulation-induced analgesia. Although thermal stimuli are used in both tests, the tail-flick response is due to spinal reflex, while the hot-plate paw-licking response is a supraspinally integrated response (Suh et al., 1992; Schmauss and Yaksh, 1984). The antinociceptive effect in the tail flick and hot-plate tests indicates that IFZN causes inhibition on both the spinal reflex and supraspinal centers (Dewey et al., 1970). Further, tail flick latency of mice treated with IFZN 30 mg/kg decreases in the presence of naloxone (opioid antagonist), which is an indicative of the involvement of the opioid receptor and the supraspinal centre.

The docking of nummularine with \( \mu \)-opioid receptor was found to be very encouraging. The docking study confirmed the interaction of cyclopeptide alkaloids with \( \mu \)-opioid receptor.

5. Conclusion

Based on results obtained in the present pharmacological study, it can be concluded that IFZN is a more potent and efficacious analgesic agent than EZN. IFZN is ten times more potent than EZN. The analgesic activity of IFZN is mediated by the peripheral as well as the central pathways mediated through the \( \mu \)-opioid receptor. The findings of present studies warrant further research on IFZN for other indications.
Acknowledgments

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References


